BILIRUBIN MONO AND BIS-DIMETHYLAMIDES. SYNTHESIS, SPECTROSCOPY **AND** SOLUTION STRUCTURES.

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 $Abstract - Bilirubin-IX\alpha bia-dimethylamide and 8- and 12-monop$ dimethylamides, and bilirubin-IIIa and XIIIa bis-dimethylamides have been synthesized and characterized spectroscopically. In deuteriochloroform, a8 deduced from El-nmr N-H resonances, the mono-dimethylamides prefer a folded, intramolecularly H-bonded conformation characteristic of the parent bilirubine. The posaiblity of similar conformations for the bie-dimethylamides is discussed.

Considerable attention has been drawn to the properties of the potentially toxic biological waste product, bilirubin-IXa (BR) because of its central role in iaundice and phototherapy.^{1,2} In particular, the three dimensional structure, as revealed by X-ray crystallography^{3,4} and nuclear magnetic resonance studies⁵ has proved to be of crucial importance in explaining its unexpected lipophilicity and the molecular mechanism of jaundice phototherapy.² One of the most interesting and important aspects of the structure of BR is its ability to exhibit intramolecular hydrogen-bonding (Figure 1).¹⁻⁵ The intramolecularly H-bonded conformation is probably the dominant species in non-H-bonding organic solvents, such as chloroform. Expectedly perhaps, the dimethyl ester of BR shows no strong tendency for intramolecular H-bonded conformations of the type shown in Figure $1.^{\circ}$ Indeed, the available evidence supports intermolecularly H-bonded dimeric structures in chloroform.³ The contrasting behavior raises the intriguing possibility that the amide8 of **BR** might exhibit conforratione either like the ester or like the acid depending on the presence of an amide N-E. In the current work, we describe the preparation of and report and interpret the spectral data for BR bie-dimethylamide $(\underline{1})$, the 8- and 12-mono-dimethylamides $(\underline{2}$ and $\underline{3}$, respectively), and the his-dimethylamides of the symmetrically substituted congenere of BR, bilirubin-IIIaand bilirubin-XIIIa.

Syntheses. The dimethylamidee were prepared by reaction of BR with dimethylamine hydrochloride using diphenylphosphoryl azide as the coupling agent in dry dimethylformamide. 8 The reaction to give the bis-dimethylamide proceeded smoothly and in high yield but was also accompanied by constitutional isomerization⁹ and afforded consequently a eeparable mixture of the bis-dimethylamides of bilirubin-III α and XIII α in addition to that of the expected IX α . With one equivalent of dimethylamine hydrochloride, **the** mono-dimethylamides could be prepared and isolated. Attempts to prepare and isolate the bis-methylamide and bis-amide of BR using methylamine hydrochloride and ammonium chloride were only partially sucessful. The reactions proceeded smoothly but the amide products exhibited such limited solubility that chromatographic separations were extremely difficult and the purity or homogeneity of material could not be ascertained.

BR: Bilirubin-IXa

- 2: **BR-8 -dimethylamide** $(X = N(CH_3)_2, Y = OH)$
- **BR-12-dimethylam** $(X=OH, Y=N(CH_3)_2)$

FIGUREl. Intramolecularly H-bonded conformations of (42,152)-bilirubin-IXa (BR) **and** its isomeric mono-dimethylamide derivatives $(2 \text{ and } 3)$.

Spectral Properties and Solution Conformation. Symmetric bilirubins (IIIa and $XIII\alpha$) and their dimethyl esters typically exhibit two N-H resonances, one assigned to the lactam and one to the pyrrole (Table 1).⁶ The non-symmetric IX α isomer and its dimethyl ester have long been known to exhibit two sets of **N-H** resonances. 5-7 All of the N-H signals are sensitive to solvent H-bonding characteristics. Thus, comparieon of the lactam and pyrrole N-H signals of the acida and esters (Table 1) in d₆-dimethylsulfoxide shows only small differences, with the lactam N-H reson**antes** falling near 10.0 ppm and the pyrrole N-H near 10.5 ppm. The various bisdimethylamide derivatives behave very much the same, and since dimethylsulfoxide is known to form H-bonds with amide 10 and pyrrole 11 N-H groups, we assume that all compounds of Table 1 are similarly H-bonded to dimethylsulfoxide and possess similar conformations. $5-8$.

In deuteriochloroform, which H-bonds less well than dimethylsulfoxide, the $N-\underline{H}$ resonances are shifted. With comparisons being made between d_6 -dimethylsulfoxide and deuteriochloroform, in the latter solvent and for bilirubin acids, the lactam N-<u>H</u> signals become relatively more deshielded ($\Delta \delta$ ~ +0.8 ppm), an indicator of intensified H-bonding, 6 and the pyrrole N-H signals become relatively more strongly

TABLE 1

a
- Data from reference 6.

 $\frac{b}{2}$ Run at 20-25°C on 2 x 10⁻³ M solutions.

shielded $(\Delta \delta \sim -1.2 \text{ ppm})$. The data (Table 1) are consistent with an intramolecularly H-bonded conformation (Figure 1) in deuteriochloroform. As with the bilirubin acid behavior, the dimethyl ester lactam N-H signals become more deshielded ($\Delta \delta$ \sim +0.8 ppm); however, the pyrrole N-H signals are less strongly shielded ($\Delta\delta$ ~ -0.3 ppm). The data here are consistent with intermolecularly H-bonded dimers in chloroform, as described earlier.⁵⁻⁷ The N-H resonances of the bis-dimethylamides (Table 1) show a qualitatively similar solvent dependence: the lactam N-H signal is more deshielded in deuteriochloroform than in $d₆$ -dimethylsulfoxide ($\Delta\delta \sim +0.5$) and the pyrrole N-H is relatively more shielded ($\Delta\delta \sim -0.7$). By way of comparison, xanthobilirubic acid, its methyl ester and dimethylamide, all pyrromethenone models that are incapable of intramolecular H-bonding and prefer intermolecularly H-bonded (pyrromethenone-to-pyrromethenone) dimeric conformation in chloroform, show a larger deshielding of the lactam N-H resonance in deuteriochloroform than in $\mathtt{d_{g}}$ -dimethylsulfoxide ($\Delta \delta$ ~ +1.5) and essentially no change in the pyrrole N-<u>H</u> resonances (ΔS \sim 0). \degree With a large pyrrole N-<u>H</u> shielding ($\Delta \delta$) taken as an indicator of intramolecular H-bonding, one is tempted to conclude that intramolecularly H-bonded conformations in chloroform may play a more important role in the structures of bilirubin bis-dimethylamides than of the dimethyl esters. In any event, the data (Table 1) show that the propionamide group plays a conformation-determining role, a role that might not be expected in an intermolecularly H-bonded dimeric structure.

Unlike the dimethyl esters and bis-dimethylamides, however, partial intramolecular H-bonding is still possible between the remaining propionic acid and opposing pyrromethenone groups (Figure 1) of the mono-dimethylamides, as with the monomethyl esters. Since an earlier analysis of the N-H chemical shifts of the monomethyl esters of bilirubins-IXa, IIIa and XIIIu concluded in favor of such intramolecular H-bonding, ¹⁴ it seemed likely that similar conformations might obtain with the mono-dimethylamides as well. Thus, if one assumes that the propionic acid groups of mono-amides 2 and 3 (and mono-esters (5 and 6) might participate in intramolecular H-bonding, the resulting conformation (Figure 1) should be expected to show one set of pyrromethenone N-H resonances at approximately the deshieldings observed in the parent 8R (Table 1). This expectation is in fact borne out in the NMR data of Table 2 in which very good agreement is found between the experimental and predicted values for the mono-amides, $\underline{cf.}$ 2: I and I', <u>3</u>: II and II', and also for the mono-esters, $\underline{cf. 4}:$ I and I', $\underline{5}:$ II and II'. We conclude, therefore, that the conformation in deuteriochloroform ia at least "half" intramolecularly H-bonded (cf. Figure 1).

The remaining pyrromethenone "half", the one not obviously involved in intramolecular H-bonding might become involved in intermolecular H-bonding of the pyrromethenone-to-pyrromethenone type characteristic of bilirubin dimethyl esters.^{6,7} Or, given the already partially folded conformation dictated by the propionic acid and opposing pyrromethenone groups (vide ante), some type of intramolecular Hbonding between the remaining propionamide and opposing pyrromethenone groups might obtain. The experimental data (Table 2) indicate a preference for the same type Of conformation in this part of the mono-dimethylamide as that exhibited by the bisdimethylamide, $c\text{f}$. 2: II and II', and 3: I and I'. The net preferred conformation of 2 and 3 in deuteriochloroform would appear to be something akin to the intramolecularly H-bonded structures of Figure 1. Consietent with this picture, the data of Table 2 for the monomethylesters are incompatible with the dimethyl ester^{6,7} or xanthobilirubic acid ester⁸ type of dimeric pyrromethenone-pyromethe none H-bonding, $\underline{\text{cf.}}$ 5: II vs II', and $\underline{\text{6}}$: I vs I'. Here, too, an even weaker intramolecular H-bonding between the propionic eater and opposing pyrromethenone groups seema likely (Figure 1).

Lactam and Pyrrole N-E Assignments in the تـ H-NMR Spectra5 ر
م of Bilirubin Mono-dimethylamides and Monomethyl Esters.-

0:
0: Assignments are indicated under the appropriate N-H. Assignments are indicated under the appropriate N-H. o:
o:

a Reported in ppm downfield from TMS for 10 -3 in CDC13 and 10 -3 in CDC13 and 10 -2 E E solutions in d₆-DMSO at 25'C. a Reported in ppm downfield from TMS for 10⁻³ M solutions in CDCl₃ and 10⁻² M solutions in d₆-DMSO at 25°C.

b Data for monomethyl esters from reference 14. - Data for monomethyl esters from reference 14.

 $\ddot{}$

 \approx Derived from data reported in Table 1, assuming one pyrromethenone behaves like the bilirubin and the other ^c Derived from data reported in Table 1, assuming one pyrromethenone behaves like the bilirubin and the other
pyrromethenone behaves like the bilirubin bis-dimethylamide or dimethyl ester. pyrromethenone behaves like the bilirubin bis-dimethylamide or dimethyl ester.

The 13 C-nmr data for the bis-dimethylamides of bilirubin-IX α , III α and XIII α are assigned and summarized in Table 3. The carbon resonances do not vary significantly, and here, but not with xanthobilirubic acid dimethylamide, 8 the amide methyl groups are non-equivalent. Non-equivalence is to be expected as the charge-separated resonance structure assumes greater importance. $8,15$ These data

^a/Me [RC-N,we t--*

thus offer further evidence for involvement of the propionamide groups in a conformationally restricted way (with neighboring pyrromethenone groups) as in an intramolecularly H-bonded structure (Figure 1). Presumably, the enhanced basicity of the propionamide oxygen strengthens intramolecular H-bonding in amides relative to esters, where the importance of the charge separated resonance structure is not as great.

In summary, we have prepared the bis-amides of bilirubin-IX α , III α and XIII α , and the 8- and 12-mono-dimethylamides of bilirubin-IXa. These have been characterized spectroscopically and have been found to exhibit a tendency toward intramolecular H-binding in chloroform solution.

EXPERIMENTAL

General: All nmr spectra were run on a JEOL FX-100 FT spectrophotometer in either d₆-dimethylsulfoxide (d₆-DMSO) (99.5% d₆) from Merck or deuteriochloroform (99.8% d_l), from Stohler. AI1 ir spectra wĕre obtained on a Perkin-Elmer model 599
instrument, and all uv-visible absorption spectra were run on a Cary 219 instrument. Nelting points were determined on a Mel-Temp capillary unit. Combuetion microanalyses were obtained from MICANAL, Tucson, AZ. Thin layer chromatography (tic) was carried out using silica gel F (M. Woelm) on analytical plates (125u) prepared from ethanol slurries with the plates being activated at 110-120' following ethanol evaporation. Preparative layer chromatography (plc) was carried out with 20 x20 cm plates having a 1 mm thick layer of silica gel F. Bilirubin was from Sigma. Dimethyl formamide, triethylamine (dried by distillation from calcium hydride) and dimethylamine hydrochloride were from Matheson, and diphenylphosphorylazide was from Aldrich.

Bilirubin-IIIa, JXa, and XIIIa Bie-dimethylamides: Bilirubin-IXa, previouely crystallized from chloroform-methanol (150.8 mg, 0.258 mmoles), freshly sublimed dfmethylamine hydrochloride (118 mg, 2.24 mmole), diphenylphosphoryl axide (0.454 ml, 0.575 mmoles) and triethylamine (0.35 ml, 0.25 gr 2.5 mmoles) were stirred in 50 ml of dried dimethylformamide in a glass pressure-bottle at 60. under Ar for 2.5 hours. The solvents were removed under vacuum (rotary evaporator) to afford a solid. Which was dissolved in chloroform. The chloroform solution was washed with 1 **N** sodium bicarbonate (2 x 25 ml) then water (50 ml) and dried (sodium sulfate) and evaporated. The resulting residue was applied to two 20 x 20 cm plc plates (1 mm1 silica gel) and developed with benzene-ethanol, 25:3 v/v. Three faint, fast moving bands (unreacted bilirubin and mono amides) were separated from the major
three bands (R_f 0.47-0.41, 0.41-0.31 and 0.31-0.21), which were themselves separated and collected to give fractions totalling 61.9 mg, 95.7 mg and 32.9 mg of the IIIa, IXa and XIIIa isomers, reepectively. The isomers were each further purified by preparative tic on bilica gel to give 31.6 mg (19%), 64.8 mg (39%) and 11.3 mg (7%) of the pure III α , IX α and XIII α isomers, respectively. The III α and IX α isomers could be crystallized from methanol: 29.8 mg of IIIa gave 19.7 mg of dark red crystals, mp 127-1320 dec. and 49.8 mg of IXa gave 42.1 mg of dark red crystals, mp 125-130', dec. The IXa isomer had uv-via (chloroform): A max 441 nm, E, 61,200, (dimethylsulfoxide): X **max** 460 nm, E, 61.899; ir (chloroform): v 3300, 3005, 1678, 1634, 1619, 1205, 1013, 925 **cm-';** 'H-nmr (deuteriochloroform): 6 ppm 1.77 (s, CH₃), 2.01 (s, 2 x CH₃), 2.3-2.8 (m, 8H), 2.88 (s, 6H), 4.00 (s, 2H), 4.96
(q, 1H, J(AB) = ~1.5 Hz, J(AX) = ~12 Hz), 5.41 (q, 1H, J(AB) = ~1.5 Hz, J(BX)=
~11.6 Hz), 5.44 (q, 1H, J(AB) ~1.5 Hz, J(BX) = ~18 Hz), 6.23 (q, 1H, J(AX) = ~12 Hz, J(BX) = ~18 Hz), 6.52 (q, 1H, J(BX) = ~18 Hz, J(AX) =
~12 Hz), 9.78 (s, 1H), 10.32 (s, 1H), 10.43 (s, 2H); ¹³C-nmr in Table 3; field desorption mass spectral molecular weight = 630.

Anal. Calcd for C₃₇H₄₆N₆O₄ (638): C, 69.59: H, 7.21: N, 13.17.

Found: C, 69.40: H, 6.85: N, 12.79.

IIIa bis-dimethylamide had mp 127-132° (dec.) uv-vis (chloroform): λ max 446 nm, ε , 69.000, (dimethylsulfoxide): 3005, **amathylaulfoxide):** λ max 464 nm, ε, 70,100; ir (chloroform): ν 3300,
1678, 1634, 1619, 1205, 1013, 925 cm ; ¹H-nmr (deuteriochloroform): δ ppm **2.02 (8,** 2 x CHJ), 2.09 (6, 2 x CHr), 2.3-2.8 (m, 8H), 2.90 (s, 2 x syn CH,- N), **2.96** (a, 2 x anti CH,-N), 3.91 (a, 2H), 5.14 (q, 2H, J(A8) = - 2.5 Hz, J(AX)= -11 Hz), 5.93 (q, 2H, J(AB) = ~2.4 Hz, J(BX) = ~18 Hz), 5.95 (s, 2B), 6.38 (q, 2H,
J(AX) = ~11 Hz, J(BX) = ~18 Hz), 9.79 (s, 2H), 10.44 (s, 2H); ¹³C-nmr in Table 3. Anal. Calcd for C_3 ,H_{b6}N₆O₄ (638): C, 69.59; H, 7.21; N, 13.17. Found: C, 69.42: **H,** 6.86: N, 12.80.

XIIIa bis-dimethylamide had 129-135° (dec.), uv-vis (chloroform): λ max 435 nm, ε , 52,900, (dimethylaulfoxide): λ max 456 nm, ε , 55,900; ¹H-nmr (deuteriochloroform): δ ppm 1.75 (s, 2 x CH₃), 2.01 (s, 2 x CH₃), 2.4-2.8 (m, 8H), 2.88 (s, 2 x syn CH₃-N), 2.91 (s, 2 x anti CH₃-N), 3.98 (s, 2H), 5.42 (q, 2H, J(AB) = ~ 2.0 Hz, J(AX) $J(AX) = -12$ Hz, $J(BX) = -17$ Hz), 10.01 (s, 2H), 10.39 (s, 2H); ¹³C-nmr in Table 3. Anal. Calcd for C₃₇H₄₆N₆O₄ (638): C, 69.59; H, 7.21; N, 13.17. Found: C, 69.43: H, 6.83: N, 12.89.

Bilirubin-IXa 8-Dimethylamide and 12-Dimethylamide. Bilirubin-IXa, previously crystallized form chloroform-methanol (201.5 mg, 0.345 mmoles), freshly sublimed dimethylamine hydrochloride (28 mg, 0.345 mmolesj, diphenylphosphoryl axide (0.224 ml, 284.6 mg, 1.035 mmoles) and triethylamine (0.144 ml, 104.5 mgr 1.035 mmoles) were combined and stirred in 50 ml of dried dimethylformamide under argon in a pressure bottle at ca 60'C for 1.5 hours. The solvents were **then removed at re**duced pressure (rotary evaporator), and the residue was dissolved in chloroform, washed with $1\,$ N $\,$ aq. sodium bicarbonate (2 x 30 ml), water (1 x 50 ml) and dried (sodium sulfate). After evaporation of the chloroform, the resulting solid was chromatographed by plc (2 plates) using chloroform-methanol-acetic acid (97:2:1, v/v/v) as eluent. The mono-amides migrated as compact bands (R $_{\rm c}$ > 0.5) that were preceded slightly by bilirubin. The yield of crude mono-amides wae 43.6 mg (20%) of 8-dimethylamide and 65.7 mg (30%) of 12-dimethylamide. These partially purified monoamides were further chromatographed twice by plc using chloroform-acetic acid 99:1 v/v) to afford 14.7 mg 97%) of pure 8-dimethyl amide (R $_{c}$ 0.2) and 31.0 mg (15%) 12-dimethylamide (R $_{e}$ 0.05). X max 446 nm, (15%) l2-dimethylamide (R, 0.05). The 8-dimethylamide isomer hãd mp 205-210° dec.
(chloroform): λ max 444 nm, ε, 51,800, (dimethylsulfoxide): λ max 460 nm, ε , 45,400; H-nmr (deuteriochloroform): δ ppm 1.92 (s, CH₃), 2.03 (s, CH₃), 2.07 (s, CH,), 2.09 (s, CH,), 2.4-2.8 (m, 8H), 2.90 (9, syn **CHI-N), 3.07 (s** anti CH,-N), 3.92 (s, 2H), 5.1-5.7 (m, 3H), 6.15 (6, 1H)r 6.0-6.6 (m, 3H), 9.18 (8, 1H)r 9.42 $(s, 1H), 10.48 (s, 1H), 10.72 (s, 1H).$

 $\frac{\texttt{Anal}}{\texttt{Anal}}$. Calcd for C₃₇H₄₆N₆O₄ (611): C, 68.72; H, 6.76; N, 11.45. Found: C, 68.52: H, 7.18; N, 11.05.

The 12-dimethylamide had mp 208-214° dec., uv-vis (chloroform): λ max 443 nm, ε,
49,800, (dimethylsulfoxide): λ max 456 nm, ε, 51,100; ¹H-nmr (deuteriochloroform): δ ppm 1.89 (s, CH₃), 1.99 (s, CH₃), 2.06 (s, CH₃), 2.08 (s, CH₃), 2.3-2.8 (m, 8H), 2.89 (s, syn CH3-N), 3.06 (s, anti CH3-N), 3.92 (s, 2H), 5.2-5.7 (m, 3H), 5.01 (s,
1H), 6.14 (s, 1H), 6.0-6.8 (m, 3H), 9.18 (s, 1H), 9.46 (s, 1H), 10.46 (s, 1H), 10.72 (s, $1H$).
Anal.

Anal. Calcd for C₃₅H₄₁N₅O₅ (611): C, 68.72; H, 6.76; N, 11.45. Found: C, 68.52; H, 7.18; N, 11.05.

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TABLE 3

13c-NMR Assignments² for Bilirubin Bis-Dimethylamides.^D

For numbering, see Figure 1; for structures, see Table 1.

k Run in d_6 -dimethylsulfoxide.